

**BIOL 419/519 Bioinformatics Research**  
**Gene Finding 2**  
**Paul Szauter – June 13, 2013**

**Introduction**

In this exercise, we will use a custom installation of the UCSC Genome Browser at GEP to view a *D. ananassae* fosmid. The UCSC Genome Browser allows a large variety of different data types to be displayed against a physical map of a fosmid or contig. We will be using the UCSC Genome Browser extensively for annotation. In this first view, we will use it to confirm and extend your analysis of the genes present on your fosmid or contig.

**1. *D. ananassae* fosmid 1475K17**

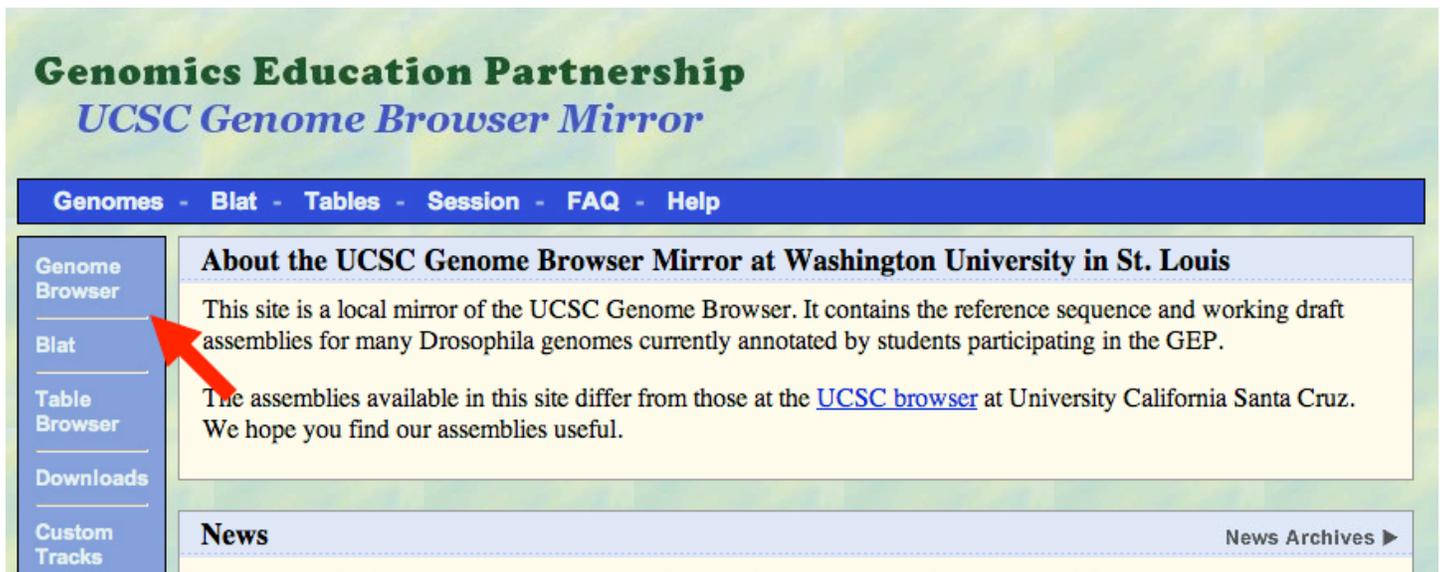
On the course website, navigate to the **Tools** page:

<http://www.discoveryandinnovation.com/bioinformatics/tools.html>

Click the link to the UCSC Genome Browser at GEP to go to:

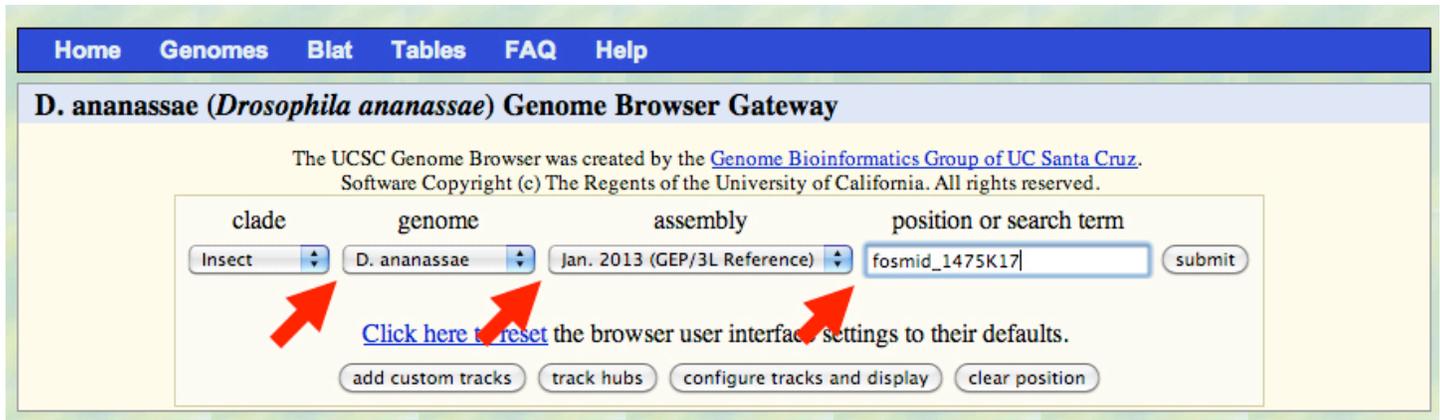
<http://gander.wustl.edu/>

In the page that appears (shown below), click the **Genome Browser** link.



In the page that appears (shown below):

1. Select **D. ananassae** as the **Genome**.
2. Select **Jan. 2013 (GEP/3L Reference)** as the **assembly**.
3. Enter **fosmid\_1475K17** as the **position or search term**.
4. Click **Submit**.



In the page that appears, there will be a visual display with many control buttons. Set the controls as described below.

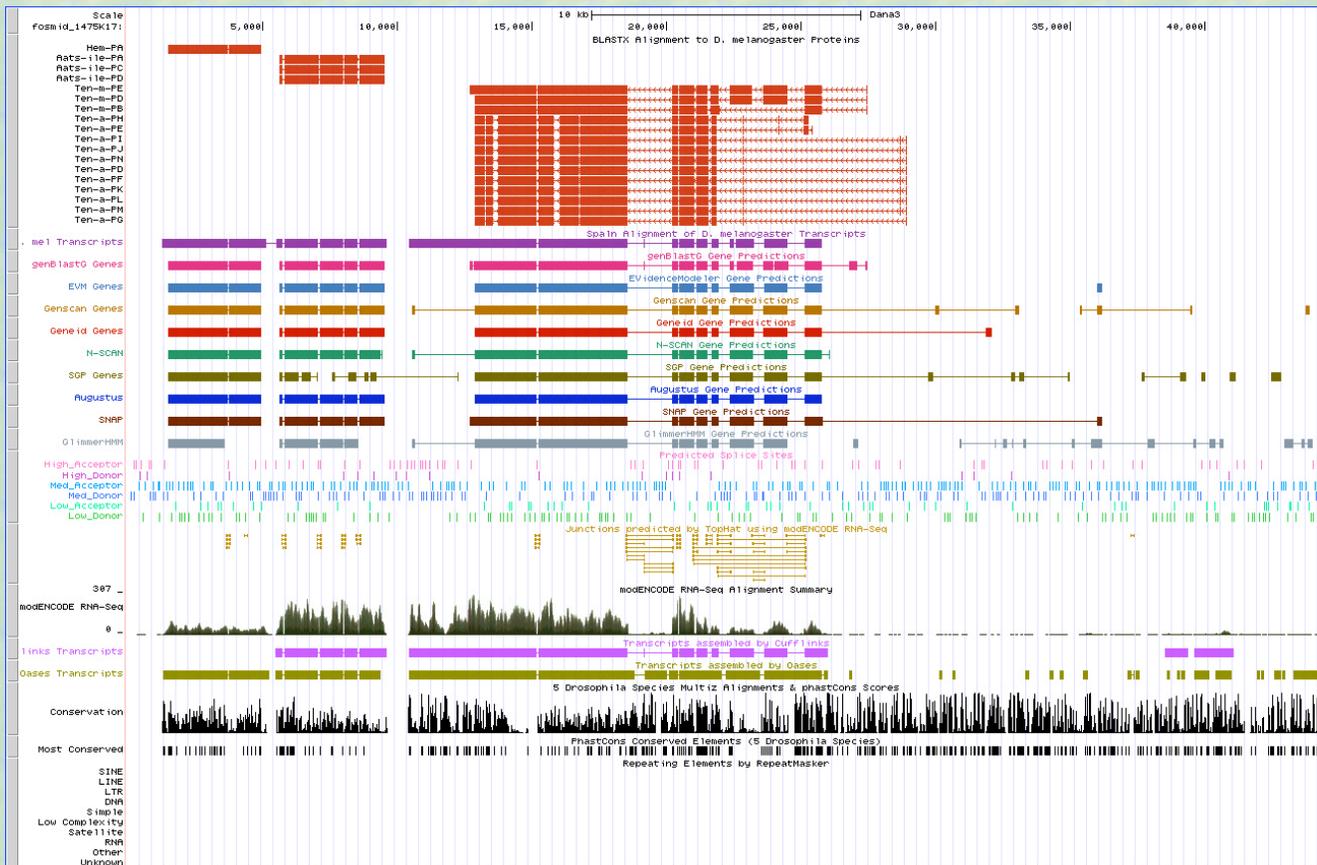
1. In **Mapping and Sequencing Tracks**, set **Base Position** to full, all other tracks to **hide**.
2. In **Genes and Gene Prediction Tracks**, set all to **dense**.
3. In **RNA Seq Tracks**, set **modENCODE TopHat junctions** to **squish**, **modENCODE RNA-Seq Summary** to **full**, **Cufflinks Transcripts** and **Oases Transcripts** to **dense**, and all others to **hide**.
4. In **Comparative Genomics**, set **Conservation** to **squish**, **Most Conserved** to **dense**, and all others to **hide**.
5. In **Variation and Repeats**, set **RepeatMasker** to **full** and **Simple Repeats** to **hide**.

Your display should now resemble the image on the next page.

# GEP UCSC Genome Browser on *D. ananassae* Jan. 2013 (GEP/3L Reference) Assembly (Dana3)

move <<< << < > >> >>> zoom in 1.5x 3x 10x base zoom out 1.5x 3x 10x

position/search fosmid\_1475K17:1-44,438 jump clear size 44,438 bp. configure



move start

< 2.0 >

Click on a feature for details. Click or drag in the base position track to zoom in. Click side bars for track options. Drag side bars or labels up or down to reorder tracks. Drag tracks left or right to new position.

move end

< 2.0 >

default tracks default order hide all add custom tracks track hubs configure reverse resize refresh

Use drop-down controls below and press refresh to alter tracks displayed. Tracks with lots of items will automatically be displayed in more compact modes.

## Mapping and Sequencing Tracks refresh

Base Position GC Percent Short Match Restr Enzymes  
 full hide hide hide

## Genes and Gene Prediction Tracks refresh

D. mel Proteins D. mel Transcripts genBlastG Genes EVM Genes Genscan Genes Geneid Genes  
 full dense dense dense dense dense  
 N-SCAN SGP Genes Augustus SNAP GlimmerHMM Predicted Splice Sites  
 dense dense dense dense dense dense

## RNA Seq Tracks refresh

modENCODE RNA-Seq modENCODE TopHat modENCODE RNA-Seq Coverage modENCODE RNA-Seq Summary Cufflinks Transcripts Oases Transcripts  
 hide squish hide full dense dense

Spliced RNA-Seq  
 hide

## Comparative Genomics refresh

Conservation Most Conserved (dm3) D. mel. Chain (dm3) D. mel. Net D. ere. Chain D. ere. Net  
 squish dense hide hide hide hide  
 D. tak. Chain D. tak. Net D. bip. Chain D. bip. Net  
 hide hide hide hide

## Variation and Repeats refresh

RepeatMasker Simple Repeats  
 full hide

refresh

**CLASS DISCUSSION:** We will discuss the meaning of the various tracks and how to use this display to confirm the results of your gene finding with BLASTX and GENSCAN.

## **2. Your fosmid or contig**

Return to the starting page and set the UCSC Genome Browser to display your *D. ananassae* fosmid or *D. biarmipes* contig. Feel free to change the track settings (always hitting refresh) to see what happens. Consider the following questions.

- a. How do the number and position of *D. melanogaster* proteins support your BLASTX results?
- b. How well do the various Gene Prediction tracks agree? How does the display compare to your BLAST analysis of GENSCAN predictions?
- c. What parts of your fosmid or contig have the greatest sequence conservation?
- d. How are the repeated sequences distributed?

Be sure to take screenshots to support your work on the genes present on your fosmid or contig.